

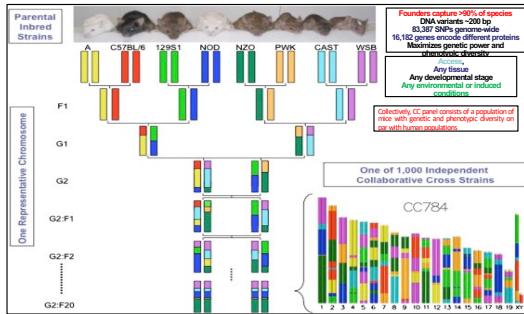
Upstream factors of micrornas identified through CC mice are involved in transcriptional regulation and neurogenesis of mNPCs

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INTRODUCTION

Collaborative cross mouse model

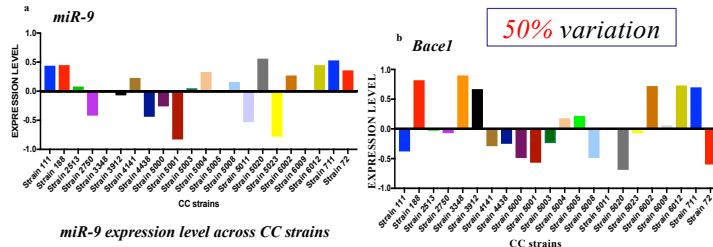


METHODS

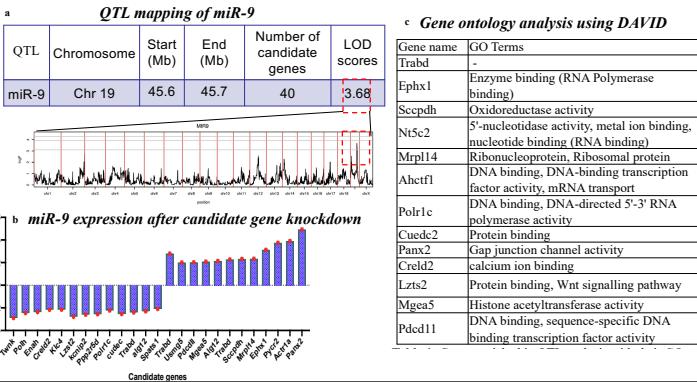
1. Validating CC mice as a model to study genetic diversity
2. Identifying upstream multiple factors through QTL mapping, siRNA knockdown, miRNA expression change and Bioinformatics
3. Functional validation of Panx2, Mgea5 and Polr1c through miR-9 pathway in neuronal differentiation
4. Identify mechanism of upstream regulation of Panx2, Mgea5 and Polr1c in miR-9 signaling using ChIP-seq, 3C, ChIP loop and Luciferase Reporter assay

RESULTS

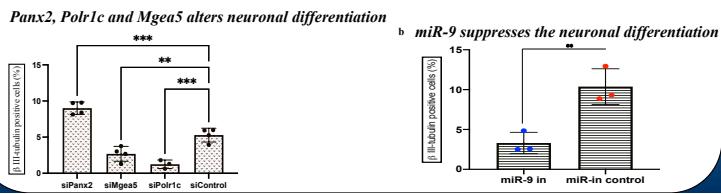
Using CC mice as a model to study genetic diversity



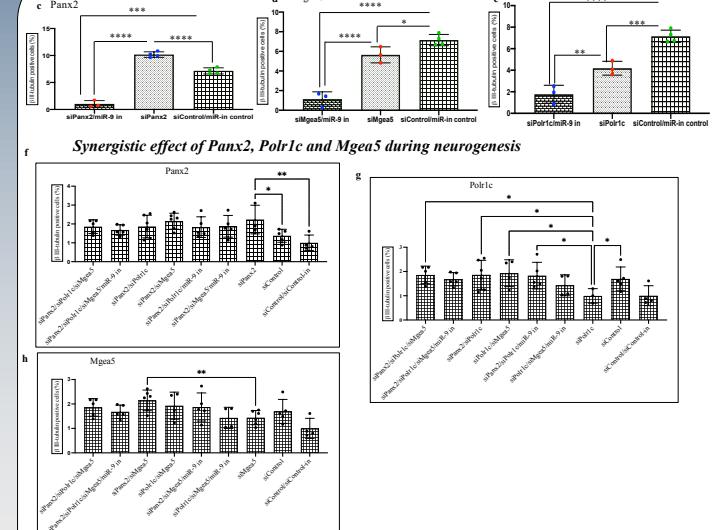
QTL mapping of the CC strains uncovers loci controlling miRNA expression



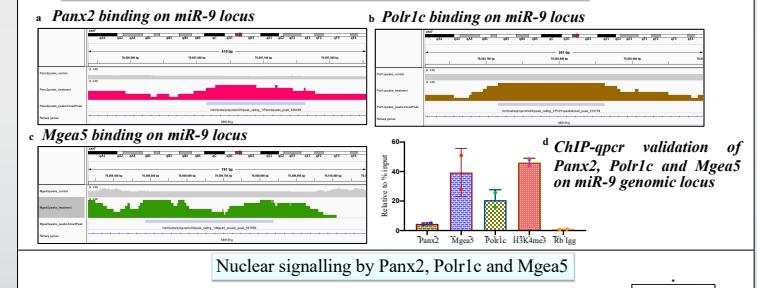
Functional validation of Panx2, Mgea5 and Polr1c in neuronal differentiation



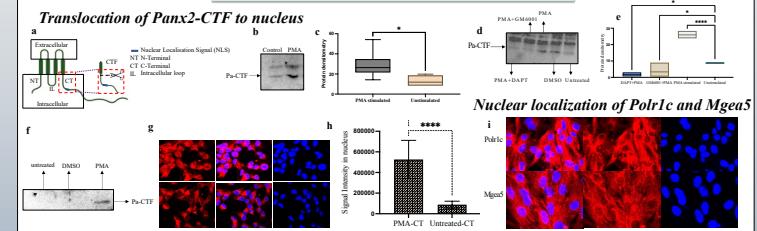
Panx2, Polr1c, Mgea5 modulates neuronal differentiation through miR-9



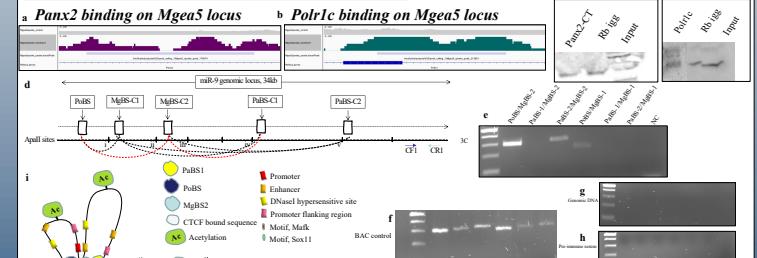
Recruitment of Panx2 Polr1c and Mgea5 on miR-9 genomic locus



Nuclear signalling by Panx2, Polr1c and Mgea5



Upstream regulation of Panx2, Mgea5 and Polr1c



CONCLUSIONS

- I. CC mice as a useful tool to study genetic diversity
- II. Panx2 has an additive effect whereas Polr1c and Mgea5 act synergistically in neuronal differentiation
- III. Panx2, Polr1c and Mgea5 is recruited to the miR-9 genomic locus
- IV. Nuclear signaling translocates Panx2-CTF to the nucleus

Panx2, Polr1c and Mgea5 forms chromatin associated loop to regulate transcription of miR-9

Novel multigenetic factors Panx2, Polr1c and Mgea5 through miR-9 signalling using CC mice for NDD research